



## High variability in the number of *E. multilocularis* eggs in cat feces collected in the field

Gérald Umhang<sup>a,\*</sup>, Matthieu Bastien<sup>b,c</sup>, Vanessa Bastid<sup>a</sup>, Marie-Lazarine Poulle<sup>c,d</sup>, Franck Boué<sup>a</sup>

<sup>a</sup> Anses, Nancy Laboratory for Rabies and Wildlife, National Reference Laboratory *Echinococcus* spp., 54220 Malzéville, France

<sup>b</sup> French Establishment for Fighting Zoonoses (ELIZ), Domaine de Pixérécourt, 54220 Malzéville, France

<sup>c</sup> University of Reims Champagne-Ardenne, SFR Cap Santé, EA 7510 ESCAPE, 51092 Reims cedex, France

<sup>d</sup> University of Reims Champagne-Ardenne, CERFE, 08240, Boult-aux-Bois, France

### ARTICLE INFO

#### Keywords:

*Echinococcus multilocularis*  
Domestic cat  
Biotic potential  
Eggs  
Zoonotic risk

### ABSTRACT

*Echinococcus multilocularis* is the causative agent of alveolar echinococcosis that is considered as the most severe parasitic disease in Europe. The contribution of cat to environmental contamination by *E. multilocularis* is generally considered as extremely low based on results of experimental infections and worm burden estimations from natural infections. However, the recent collection of numerous cat feces from kitchen gardens in high endemic areas and the detection of *E. multilocularis* DNA in a significant number of these feces raise the question of the risk of human transmission from cats. This study aimed to provide a quantitative estimation of *E. multilocularis* eggs in feces from naturally infected cats. A field sampling conducted in 192 kitchen gardens during a joint study led to the collection and analysis of 597 cat feces, among them 7 (1.2%) yielded positive results for *E. multilocularis* real-time PCR. The entire pellets obtained after homogenization, filtration and centrifugation of a 5 g-sample for each of these 7 feces were examined under a stereoscopic microscope. After assessing their number, 20 taeniid eggs were individually isolated and specifically identified by real-time PCR. Morphologically mature *E. multilocularis* eggs were identified in 4 samples and the counting of 4 to 43 *E. multilocularis* eggs per gram in these samples, i.e. 62 to 2331 eggs per feces when the total mass of the feces is considered. The number of eggs counted in 2 feces suggests a biotic potential of some naturally infected cats that largely exceed the previous experimental estimations.

The cestode *Echinococcus multilocularis* is the causative agent of alveolar echinococcosis that is considered as the most severe parasitic disease in Europe [1]. The lifecycle in Europe is essentially sylvatic and based on predation of small rodents by wild canids, mainly red fox (*Vulpes vulpes*). The rodent intermediate hosts are infected after ingestion of microscopic eggs, which are capable of surviving in the environment during months due to their resistant outer envelope [2]. After reaching the liver, the oncospheres will develop into metacestodes leading to production of protoscoleces in around two to four months. The viable protoscoleces are infectious for the definitive host when it ingests the parasitized prey. The intestinal development of protoscoleces into gravid worms with egg production will generally take between 28 and 35 days after infection. The total number of eggs excreted during one infection (i.e. biotic potential) is variable, notably according to host species [3,4]. A comparative study of experimental infection by

*E. multilocularis* in definitive hosts described a 500-fold higher biotic potential for red foxes, raccoon dogs (*Nyctereutes procyonoides*) and dogs (*Canis familiaris*) than for domestic cats (*Felis catus*) [3]. Very low worm burden (3 to 30–50 worms) were generally found in the intestine of naturally infected cats [5,6]. A higher worm burden (7040 worms) was described by Umhang et al. [7], but consisted of only immature worms, and only a small proportion of them are assumed to reach the fully mature adults to release eggs. Both because of its low worm burdens and its excretion of a very small number of eggs of unproven infectivity, the cat is generally considered to play an insignificant role in the transmission of *E. multilocularis* [3]. However, the evidence of frequent consumption by cat of *E. multilocularis* intermediate host species [8], the detection of DNA of this parasite in many cat feces [7,9–13] and the high density of cat feces in kitchen gardens in endemic areas [14] raise concerns about the zoonotic risk associated to cat infections.

\* Corresponding author.

E-mail address: [gerald.umhang@anses.fr](mailto:gerald.umhang@anses.fr) (G. Umhang).

<https://doi.org/10.1016/j.parint.2022.102583>

Received 17 January 2022; Received in revised form 2 April 2022; Accepted 2 April 2022

Available online 6 April 2022

1383-5769/© 2022 Elsevier B.V. All rights reserved.

Information on the biotic potential of naturally infected cats is needed to assess the epidemiological status of this host with respect to *E. multilocularis*.

The aim of this study was to estimate the total number of *E. multilocularis* eggs in feces of naturally infected cats from endemic areas. This quantitative estimation was then compared to that obtained from experimental infections to discuss the contribution of the cat to environmental contamination by *E. multilocularis*. A field sampling was conducted from January 2014 to December 2015 by Bastien et al. [15] in 192 kitchen gardens located in high endemic regions for *E. multilocularis* in northeastern France. It resulted to the collect of 1016 carnivore feces. Real-time coproPCR assays identified 597 of these feces as being from cats and detected *E. multilocularis* DNA in seven of them [9]. All the seven feces originated from different kitchen gardens. These feces were stored frozen since collection including one week at  $-80^{\circ}\text{C}$  for decontamination. Five grams were homogenized in 50 ml of distilled water and then filtered through a 120  $\mu\text{m}$  nylon mesh. After centrifugation, the supernatant was discarded and the pellets completed again with 50 ml of distilled water before to be homogenized. These steps were repeated at least two times to clear the pellets. The entire pellets were then examined under a stereoscopic microscope (x200) and all the taeniid eggs observed during this examination were counted. When present, 20 taeniid eggs were individually isolated for each fecal sample in order to be submitted to individual DNA extraction (Nucleospin Tissue, Macherey-Nagel). A real-time PCR assay was performed to detect DNA of *E. multilocularis* with sequencing confirmation [16]. Five microliters of DNA were used in a final volume of 20  $\mu\text{l}$  using Maxima Probe master mix and an internal control. All samples were tested in duplicate. The real-time PCR assays were realized using a Mx3005P thermocycler (Agilent) with an initial step of  $95^{\circ}\text{C}$  for 10 min and 45 cycles of  $95^{\circ}\text{C}$  for 15 s and  $60^{\circ}\text{C}$  for 60s. When a Ct value was obtained for at least one duplicate, sequencing was realized from the PCR products to confirm identification of *E. multilocularis*. If no DNA of *E. multilocularis* was detected, the DNA samples were submitted to an endpoint PCR using primer Cest4-Cest5 in order to identify the Taeniidae species involved after sequencing [17]. In absence of detection for the two PCR assays (*E. multilocularis* or Taeniidae species), the eggs were excluded for the calculation of the number of eggs per gram (epg). We used the ratio of *E. multilocularis* eggs versus other Taeniidae species obtained from the successful amplification among the 20 eggs isolated to estimate the number of *E. multilocularis* eggs per gram and calculate the total number of these eggs in the cat's feces.

No taeniid eggs were observed after filtration for 2/7 fecal samples from which *E. multilocularis* DNA was detected. Nevertheless, *Toxocora* sp. eggs were present for these two fecal samples confirming a correct flotation. Identification of *E. multilocularis* eggs was obtained for 4/5 samples with taeniid eggs, including one with coinfection by *Hydatigera taeniaeformis*. In the only fecal sample free of *E. multilocularis* eggs among these five samples, taeniid eggs were all corresponding to the cat tapeworm *Hydatigera taeniaeformis* (Table 1). The lowest average Cq values from fecal samples were obtained from the four samples with *E. multilocularis* eggs (30.6 to 35.8) compared to the three without eggs (36.9 to 39.2). The microscopic observation revealed morphologically mature *E. multilocularis* eggs with the presence of embryophores surrounding the oncosphere in the four cases. When present, the number of *E. multilocularis* eggs per gram (epg) was ranging from 4 to 43. Considering the total mass of each fecal sample, 62 to 2331 *E. multilocularis* eggs per fecal sample were estimated.

Our finding of a slightly lower amount of DNA in feces without eggs than in feces with eggs confirms that a substantial quantity of the coproDNA from the parasite may not originated from eggs but rather from worm tissue cells. In addition, parasitic DNA detected in definitive host (i.e. cat) feces can also come from ingestion of infected prey if the feces were produced within a few days of that ingestion as it has been demonstrated for *Toxoplasma gondii*, [18]. Worm tissues or parasitic DNA from infected prey can explain alone the detection of

**Table 1**  
Information and coprologic results of the seven cat feces with detection of *E. multilocularis* DNA by copro real-time PCR.

Sample ID	Village	Kitchen garden ID	Collection date	Total weight of the fecal sample (g)	Average Cq values of <i>E. multilocularis</i> qPCR	Presence of taeniid eggs	Proportion of <i>E. multilocularis</i> among taeniid eggs	Number of <i>E. multilocularis</i> eggs/g	Total number of <i>E. multilocularis</i> eggs/fecal sample	Proportion of <i>H. taeniaeformis</i> among taeniid eggs	Number of <i>H. taeniaeformis</i> eggs/g
10,426	Faxe	FAX08	07-11-2014	54.2	30.6	yes	18/20	43	2331	0/20	0
10,471	Grandpré	GRA07	23-01-2015	36.3	35.5	yes	6/20	33	1198	2/20	22
10,371	Lorry-mardigny	LOR09	18-12-2014	8.4	34.8	yes	15/20	14	118	0/20	0
10,459	Boult-aux-Bois	BAB13	21-01-2015	15.4	35.8	yes	15/20	4	62	0/20	0
10,550	Avricourt	AVR12	10-03-2015	28.6	36.9	yes	0/20	0	0	15/20	10
10,390	Avricourt	AVR08	18-12-2014	52.5	37	no	/	/	/	/	/
11,885	Faxe	FAX08	16-12-2015	42.4	39.2*	no	/	/	/	/	/

\* Cq value obtained for only one duplicate.

*E. multilocularis* in the three feces where no eggs were observed. Regarding feces with *E. multilocularis* eggs, the DNA from these eggs comes in addition to the worm tissue source facilitating the molecular detection. A significant difference between distribution of Cq values from fox feces with or without eggs was also previously reported by Da Silva et al. [19].

Based on an experimental infection with 20,000 protozoa, the mean biotic potential of cats was estimated of 573 eggs excreted during an entire patent period estimated at 13 days in cats [3]. However, a highest daily eggs excretion of 856 eggs at 28 dpi was described for one cat in the same study. By comparison, the estimation of the number of eggs excreted in two fecal samples from our field sampling is 2 to 4-fold higher than this estimated biotic potential. The availability of only one fecal sample for each infection prevent us to assess duration of the patent periods in these natural infections, as well as the total number of eggs excreted in the course of the whole patent period. However, one can speculate that in each infection related to these two cases, several others feces from the same cats during the patent period will also containing eggs, potentially even more than described in the analyzed feces. Therefore, our data suggest that the biotic potential of some cats may be largely higher than expected even if the null or low eggs production associated with the three others positive fecal samples are in accordance to the very low biotic potential generally described for the species. The estimation of 440 eggs per gram reported for a cat from Hokkaido Island, Japan [20] which is largely (i.e. 10 fold) exceeding our highest estimations argues to excretion of large quantities of *E. multilocularis* eggs by cats from natural infections not being a very rare event. The high excretion of eggs by some cats may explain the detection of *E. multilocularis* DNA from soil samples of kitchen gardens where no fox feces were observed, including fenced kitchen gardens [9].

In a general way, natural infection in cats by *E. multilocularis* is generally restricted to a molecular diagnostic, and the number of eggs is rarely estimated when the presence of *E. multilocularis* eggs is confirmed resulting to scarce data related to the biotic potential. The data obtained from experimental infections are very informative regarding the global epidemiological situations but may underlooked some specific situations. Combination of different cat species, quantities of protozoa, *E. multilocularis* strains, ecological contexts and immunological exposures will results to a large variety of possibilities regarding the success of natural infections. The strong predation of rodents by cats can notably increase the chance of repeated infections and raise the need to assess the potential development of an immunity decreasing the worm burden and patent period, as described for foxes and dogs [21,22]. It also has to be noted that the high number of eggs excreted by infected cats in this study remains very inferior to the biotic potential of canids, as notably red foxes and dogs can excrete 279,910 and 346,473 eggs per individual, respectively [3]. Furthermore, the existence of a zoonotic risk associated to *E. multilocularis* eggs excreted via cat feces is currently not confirmed. Eggs obtained from feces of a cat experimentally infected by a North American strain of *E. multilocularis* has developed to metacestode after ingestion by a brown lemming [23] but those obtained after experimental infection of cats by a European strain has not been successful to infect mice, unlike those obtained from foxes, dogs and raccoon dog feces [3].

In conclusion, our data show that some cat feces may contain high numbers of *E. multilocularis* eggs, although the overall contribution of cats to environmental contamination by this parasite is probably extremely low compared to that of foxes. The presence of high numbers of *E. multilocularis* eggs in some cat feces reinforce the need to test the infectivity of these eggs obtained in natural conditions, which has not yet been reported. Specific protocols coupling rapid molecular diagnostic from decontaminated part of the cat fecal sample potentially followed by isolation of taeniid eggs are required. In absence of current in vitro test for evaluation of the infectivity, in vivo assays in rodents remains the only way to confirm the infectivity, and then a zoonotic potential.

## Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

## Declaration of Competing Interest

The authors declare no competing interests in association with this study.

## References

- [1] T. Romig, *Echinococcus multilocularis* in Europe—state of the art, Vet. Res. Commun. 33 (Suppl. 1) (2009) 31–34.
- [2] P. Veit, B. Bilger, V. Schäd, J. Schäfer, W. Frank, R. Lucius, Influence of environmental factors on the infectivity of *Echinococcus multilocularis* eggs, Parasitology 110 (Pt 1) (1995) 79–86.
- [3] C.M. Kapel, P.R. Torgerson, R.C. Thompson, P. Deplazes, Reproductive potential of *Echinococcus multilocularis* in experimentally infected foxes, dogs, raccoon dogs and cats, Int. J. Parasitol. 36 (1) (2006) 79–86.
- [4] R.C.A. Thompson, C.M. Kapel, R.P. Hobbs, P. Deplazes, Comparative development of *Echinococcus multilocularis* in its definitive hosts, Parasitology 132 (Pt 5) (2006) 709–716.
- [5] A.F. Petavy, F. Tenora, S. Deblock, V. Sergent, *Echinococcus multilocularis* in domestic cats in France. A potential risk factor for alveolar hydatid disease contamination in humans, Vet. Parasitol. 87 (2–3) (2000) 151–156.
- [6] G. Wobeser, The occurrence of *Echinococcus multilocularis* (Leukart, 1863) in cats near Saskatoon, Saskatchewan, Can. Vet. J. 12 (3) (1971) 65–68.
- [7] G. Umhang, M.-A. Forin-Wiart, V. Hormaz, C. Caillot, J.-M. Boucher, M.-L. Poulle, F. Boué, *Echinococcus multilocularis* detection in the intestines and feces of free-ranging domestic cats (*Felis s. catus*) and European wildcats (*Felis s. silvestris*) from northeastern France, Vet. Parasitol. 214 (1–2) (2015) 75–79.
- [8] M.A. Forin-Wiart, M.L. Poulle, S. Piry, J.F. Cosson, C. Larose, M. Galan, Evaluating metabarcoding to analyse diet composition of species foraging in anthropogenic landscapes using ion torrent and Illumina sequencing, Sci. Rep. 8 (1) (2018) 17091.
- [9] A.M. Da Silva, M. Bastien, G. Umhang, F. Boué, V. Bastid, J.-M. Boucher, C. Caillot, C.P. de Garam, C. Renault, M. Faisse, S. Courquet, V. Scalabrino, L. Millon, J. Knapp, M.-L. Poulle, Soil contamination by *Echinococcus multilocularis* in rural and urban vegetable gardens in relation to fox, cat and dog faecal deposits, Parasite 28 (2021) 74.
- [10] J. Karamon, J. Sroka, J. Dąbrowska, E. Bilska-Zajac, J. Zdybel, M. Kochanowski, M. Różycki, T. Cencek, First report of *Echinococcus multilocularis* in cats in Poland: a monitoring study in cats and dogs from a rural area and animal shelter in a highly endemic region, Parasit. Vectors 12 (1) (2019) 313.
- [11] J. Knapp, B. Combes, G. Umhang, S. Aknouche, L. Millon, Could the domestic cat play a significant role in the transmission of *Echinococcus multilocularis*? A study based on qPCR analysis of cat feces in a rural area in France, Parasite 23 (2016) 42.
- [12] J. Knapp, P. Giraudoux, B. Combes, G. Umhang, F. Boué, Z. Said-Ali, S. Aknouche, C. Garcia, M. Vacheyrou, A. Laboisière, V. Raton, S. Comte, S. Favier, J. M. Demerson, C. Caillot, L. Millon, F. Raoul, Rural and urban distribution of wild and domestic carnivore stools in the context of *Echinococcus multilocularis* environmental exposure, Int. J. Parasitol. 48 (12) (2018) 937–946.
- [13] M.-L. Poulle, M. Bastien, Y. Richard, E. Josse-Dupuis, D. Aubert, I. Villena, J. Knapp, Detection of *Echinococcus multilocularis* and other foodborne parasites in fox, cat and dog faeces collected in kitchen gardens in a highly endemic area for alveolar echinococcosis, Parasite 24 (2017).
- [14] M. Bastien, A. Vaniscotte, B. Combes, G. Umhang, E. Germain, V. Gouley, A. Pierlet, T. Quintaine, M.-A. Forin-Wiart, I. Villena, D. Aubert, F. Boué, M.-L. Poulle, High density of fox and cat faeces in kitchen gardens and resulting rodent exposure to *Echinococcus multilocularis* and *Toxoplasma gondii*, Folia Parasitol. 65 (1) (2018).
- [15] M. Bastien, A. Vaniscotte, B. Combes, G. Umhang, V. Raton, E. Germain, I. Villena, D. Aubert, F. Boué, M.-L. Poulle, Identifying drivers of fox and cat faecal deposits in kitchen gardens in order to evaluate measures for reducing contamination of fresh fruit and vegetables, Food Waterborne Parasitol. 14 (2019), e00034.
- [16] J. Knapp, G. Umhang, M.-L. Poulle, L. Millon, Development of a real-time PCR for a sensitive one-step copro-diagnosis allowing both the identification of carnivore feces and the detection of *Toxocara spp.* and *Echinococcus multilocularis*, Appl. Environ. Microbiol. 82 (10) (2016) 2950–2958.
- [17] D. Trachsel, P. Deplazes, A. Mathis, Identification of taeniid eggs in the faeces from carnivores based on multiplex PCR using targets in mitochondrial DNA, Parasitology 134 (6) (2007) 911–920.
- [18] M.-L. Poulle, M.-A. Forin-Wiart, É. Josse-Dupuis, I. Villena, D. Aubert, Detection of *Toxoplasma gondii* DNA by qPCR in the feces of a cat that recently ingested infected prey does not necessarily imply oocyst shedding, Parasite 23 (2016) 29.
- [19] A.M. Da Silva, S. Courquet, F. Raoul, D. Rieffel, P. Giraudoux, L. Millon, J. Knapp, Assessment of the exposure to *Echinococcus multilocularis* associated with carnivore faeces using real-time quantitative PCR and flotation technique assays, Int. J. Parasitol. 50 (14) (2020) 1195–1204.
- [20] N. Nonaka, H. Hirokawa, T. Inoue, R. Nakao, S. Ganzorig, F. Kobayashi, M. Inagaki, K. Egoshi, M. Kamiya, Y. Oku, The first instance of a cat excreting *Echinococcus multilocularis* eggs in Japan, Parasitol. Int. 57 (4) (2008) 519–520.

- [21] M.N. Al-Sabi, C.M. Kapel, P. Webster, P. Deplazes, Reduced egg production of *Echinococcus multilocularis* in experimentally infected and re-infected red foxes (*Vulpes vulpes*), *Vet. Parasitol.* 155 (1–2) (2008) 59–66.
- [22] H. Kouguchi, T. Irie, J. Matsumoto, R. Nakao, Y. Sugano, Y. Oku, K. Yagi, The timing of worm exclusion in dogs repeatedly infected with the cestode *Echinococcus multilocularis*, *J. Helminthol.* 90 (6) (2016) 766–772.
- [23] R.L. Rausch, S.H. Richards, Observations on parasite-host relationships of *Echinococcus multilocularis* Leuckart, 1863, in North Dakota, *Can. J. Zool.* 49 (10) (1971) 1317–1330.